# **Histamine H3 Receptor Function and Ligands: Recent Developments**

Mahaveer Singh and Hemant R. Jadhav\*

*Department of Pharmacy, Birla Institute of Technology and Sciences Pilani, Pilani Campus, Vidya Vihar – 333031, Rajasthan, India* 

**Abstract:** Histamine  $H_3$  receptors are found mostly in central nervous system involved in the regulation of release of various neurotransmitters in brain. They have been implicated in diverse potential therapeutic applications such as sleepwake disorders, attention-deficient hyperactivity disorder, epilepsy, cognitive impairment and obesity. This review is an attempt to elucidate the function of  $H_3$  receptors and their role in various CNS disorders. Also, it is aimed at collating the information on efforts of various medicinal chemists to synthesize the  $H_3$  receptor agonists and antagonists in single article.

**Keywords:** Epilepsy, H3 Receptors, Obesity, H3 Receptor Antagonists.

# **INTRODUCTION**

 Histamine (HA) plays a key role in functioning of central and peripheral tissues. It is one of the important local chemical mediators and neurotransmitters found in human body [1]. In the central nervous system (CNS), histamine is stored in vesicles of histaminergic neurons, which are located only in tuberomammilary nucleus of hypothalamus [2]. It is produced by decarboxyalation of histidine and it has wide range of physiological and pathophysiological functions in body [3, 4]. Histamine receptor has four subtypes, out of which, antagonists of  $H_1$  receptors  $(H_1R)$ and  $H_2$  receptors ( $H_2R$ ) are widely used for treatment of allergy and ulcer, respectively  $[5]$ . H<sub>3</sub> histamine receptors  $(H_3R)$  are found in CNS and to a lesser extent peripheral nervous system (PNS). These receptors, when occupied by histamine, cause decrease in release of neurotransmitters such as histamine, Gamma Amino Butyric Acid (GABA), Acetyl Choline (ACh), Norepinephrine (NE), Serotonin (5HT), and Dopamine (DA).  $H_4$  histamine receptors are found primarily in the basophils, thymus, small intestine, spleen, colon, bone marrow and play a role in chemotaxis [6].

# **H3 RECEPTOR AND ITS FUNCTION IN CNS**

Arrang *et al.* in 1983 recognized H<sub>3</sub>R and its role in Central Nervous System (CNS) as presynaptic auto receptor. Later, it was realized that  $H_3R$  is hetero as well as autoreceptor. Activation of H3R reduces the release and deactivation increases the release of various neurotransmitters in brain (Fig. 1) [7, 8]. Presynaptic neuronal control via  $H_3$ receptors is an important regulatory mechanism of HA mediated neurotransmission [9]. Histaminergic neurons are exclusively found in tuberomammilary nucleus of posterior hypothalamus and control all major areas of brain which are involved in sleep, wakefulness, cognition, transmission and various CNS functions [10].

The cloning of human  $H_3R$  was done in 1999 by Lovenberg and co-workers [11]. The overall sequence homology of the H<sub>3</sub>R to H<sub>1</sub>R and H<sub>2</sub>R is only 22% and 20%, respectively [12]. The gene of  $H_3R$  has complex structure, so a large number of  $H_3R$  isoforms exist. This is possible because of alternative splicing of  $H_3R$  mRNA. The  $H_3R$  gene consists of three exons and two introns. So far, at least 20 isoforms of the human  $H_3R$  have been identified on the basis of detection of varying mRNAs, but their regional expression and function remains largely unknown. The full-length  $H_3R$ (445 amino acids) is currently the best characterized isoform [13]. Most splice variants have deletions in the e3 loop, an important region involved in G protein coupling. In recombinant systems, it has been shown that these isoforms have altered signaling properties compared to the full-length receptor [14].

Stimulation of  $H_3R$  gives negative feedback about the synthesis of HA via adenyalate cyclase (AC) inhibition. AC catalyzes the formation of the second messenger cyclic adenosine mono phosphate (cAMP) and reduction of cAMP levels leads to inhibition of histidine decarboxylase (HDC), a key enzyme in histamine synthesis. Thus, activation of  $H_3R$ leads to decrease in histamine synthesis in presynaptic histaminergic neurons(Fig. **2**) [15].

The released HA, via presynaptic  $H_3R$  decreases the further release of HA and also inhibits release of various other neurotransmitters such as GABA, DA, NE, 5HT etc. Thus,  $H_3R$  helps in inhibitory control of histaminergic neurons via HA, as well as of the postsynaptic histamine receptors.

<sup>\*</sup>Address correspondence to this author at the Department of Pharmacy, Birla Institute of Technology and Sciences Pilani, Pilani Campus, Vidya Vihar, Pilani-333031, Rajasthan, India; Tel: +91-94146-48696; Fax: +91- 1596-244183; E-mails: hemantrj@gmail.com; hemantrj@bits-pilani.ac.in



Fig. (1). H<sub>3</sub>R Auto- and hetero- receptor function in central nervous system.



Fig. (2). H<sub>3</sub>R-mediated signal transduction – histamine synthesis and release. H<sub>3</sub>R, histamine H<sub>3</sub> receptors: AC, adenylate cyclase; PKA, protein kinase A; HDC, histidine decarboxylase; VACC, voltage-activated calcium channels; CaMKII, calmodulin kinase II.

#### **H3 RECEPTORS AND ROLE IN EPILEPSY**

 Almost 50 years ago first clinical indication suggesting the involvement of central HA mediated neurotransmission in epilepsy was reported  $[16, 17]$ . H<sub>1</sub> receptor activation or modulation of CNS HA levels by L-histidine loading and inhibition of metabolism of HA in rodents indicates that HA can be an endogenous antiepileptic [18]. The role of  $H_1$ receptors in epilepsy was further supported by an increased  $H_1$  receptor density (10 to 50 %) by focusing on epileptic discharges in the temporal neo cortex of patients with complex partial seizures and also by observation that in several young  $children, H<sub>1</sub> receptor antagonists often produced convulsions$ [19, 20].

 The involvement of HA in epileptic seizures has long been recognized based on the proconvulsant properties of histamine antagonists [21]. Histamine receptor ligands, which increase histamine levels, such as histidine and histamine N-methyl transferase (HNMT) inhibitors (eg. metoprine) reduce seizures in epileptic patients via  $H_1$  mediated excitation [22-24].  $H_3R$  antagonists such as thioperamide, clobenpropit, AQ0145 also decrease the seizure susceptibility of electrically induced convulsions in mice [25]. These compounds act by increasing endogenously released HA in the brain [26, 27]. The anticonvulsant effect of these compounds can be antagonized by giving  $H_3R$  agonists (immepip) or  $H_1$ receptor antagonist (mepyramine). These research findings suggest that  $H_3R$  antagonists could represent a new way in the development of antiepileptic drugs.

 H3R modulates release of various neurotransmitters such as NE, ACh, DA, 5HT and GABA.  $H_3R$  activation reduces release of NE in rat hypothalamus. Experiments have shown that  $H_3R$  activation decreases the release of ACh in rat cortex. In mouse stratial tissue, DA release is also reduced by histamine receptor activation. DA induced release of GABA is also inhibited by  $H_3R$  activation and presynaptic H3R mediated release of glutamate has been reported.

### **H3R and GABA Release**

The  $H_3R$  antagonist clobenpropit enhances release of GABA from rat hypothalamus [28, 29]. GABA is an inhibitory neurotransmitter, which balances the excitatory activity in synapses in CNS. GABA, as neurotransmitter, slows electrical transmission between the nerve cells and helps in balancing

of nerve impulse. Low levels of GABA in the body have been linked to epilepsy and an increased risk for seizure. A number of the drugs used to treat epilepsy stimulate production of GABA [30].

### **H3R and ACh Release**

 The relation of ACh with seizure has not been well established. ACh injection to the epileptic patient reduces the frequency of seizure. However, role of ACh in the treatment of epilepsy has been reported with different results [31]. ACh release was reduced by H3R agonists like Imetit, alpha methyl histamine, etc. This reduced release of ACh was again enhanced by H3R antagonists such as thioperamide. H3R mediated ACh release from rat basolateral amygdale and hippocampal region has also been reported [32]. Histamine receptor agonist alpha methyl histamine and imepip directly administered to rat basolateral amygdale triggered spontaneous release of ACh and administration of  $H_3R$ antagonists eg. thioperamide, ciproxifan increased the release of acetylcholine via  $H_3R$  mediated neurotransmission [33, 34]. In the medial septum–diagonal band of freely moving rats,  $H_3R$  antagonists thioperamide and ciproxifan increased the release of hippocampal ACh, whereas  $R-\alpha$ methyl histamine, an  $H_3R$  agonist, produced the opposite effect [35]. So these findings suggest that  $H_3R$  antagonists have a profound effect on release of ACh and could be useful for treatment for epilepsy.

### **H3R and 5HT Release**

 The substantia nigra pars reticulata (SNr) plays an important role in basal ganglia function. Substantia nigra pars reticulata is characterized by rich aminergic input that includes dopaminergic dendrites and axons containing 5HT or HA. The role of HA in the SNr is motor control via histamine H<sub>3</sub> receptors, the mechanism remains far from elucidated. Selective  $H_3R$  agonists R- $\alpha$ -methyl-histamine or immepip can inhibit the 5-HT release by up to 60% [36]. This inhibition can be prevented by the  $H_3R$  antagonists like thioperamide [37]. So  $H_3R$  antagonists can increase the serotonin release and gives a hope for treatment of epilepsy.

# **ROLE OF H3R IN OTHER DISORDERS**

# **Obesity**

 Hypothalamic histaminergic neurons interfere with orexigenic pathways and feeding behavior. Therefore, studies suggest that  $H_3R$  antagonists can be used for the treatment of obesity [38]. Elevated levels of central histamine reduce food intake and body weight in rodents [39]. HA activates postsynaptic  $H_1$  receptors in the ventro medial nuclei of the hypothalamus to suppress appetite and HA containing neurons in the hypothalamus participate in the endogenous suppression of food intake. In addition, there are evidences that  $H_3R$  have been found in ventro medial nuclei region [40]. Feeding is suppressed in rat by Intra-cerebroventricular injections of HA, whereas the use of  $H_3R$  antagonists such as thioperamide also supressed feeding and depletion of endogenous HA exerts feeding response [41-44]. Dysfunctions in HA mediated neurotransmission have also been identified in obese zucker rats [45].

# **Sleep and Wakefulness**

 The histaminergic system performs a major role in the maintenance of sleep and wakefulness. The presence of HA containing cells in the tuberomammillary nucleus of the posterior hypothalamus has been identified. This area is mainly involved in the regulation of wakefulness and projections of these histaminergic neurons to the cerebral cortex suggest a role of HA in the modulation of the arousal state and sleep-wake cycle. Lesions of posterior hypothalamus produce sleep in rats, cats and monkeys [46]. The histaminergic neurotransmission promotes waking and alertness. Wake promoting activity of  $H_3R$  inverse agonists has been observed in cats, mice, rats, and guinea pigs [47-49]. Modulation of histamine levels mediated by  $H_3R$ agonists results in an increase of the slow-wave sleep in rats [50, 51]. Neurochemical and electrophysiological studies indicate that the activity of histaminergic neurons is maximal during periods of wakefulness and is depressed by barbiturates and other hypnotics [52, 53]. Moreover, in some *in vivo* studies, release of histamine in the rat hypothalamus shows a circadian rhythm, with higher histamine release in periods with high locomotors activity. On the contrary, systemic application of  $H_3R$  antagonists produce increased wakefulness (decreases in rapid eye movement (REM) and slow-wave sleep) and increased locomotion.

# **Cognition and Memory Processes**

 Dysfunction of ACh -mediated neurotransmission is one of the factors responsible for cognitive decline associated with ageing and Alzheimer's disease. Histaminergic modulation of hippocampal ACh release has been reported [54]. However, changes typical of ageing and Alzheimer's disease with alterations of other neurotransmitter systems, including HA have also been reported [55, 56]. There is direct evidence that histaminergic neurotransmission plays an important role in learning and memory.

 In one of the studies, it was found that HA content was significantly reduced in the hypothalamus, hippocampus and temporal cortex of Alzheimer's brains. HA levels in other cortical areas, putamen and substantia nigra were not much different. Further, administration of HA enhanced cognitive performance of rats in an active avoidance task, while  $H_1R$ antagonists impaired memory retention [57]. Neurochemical studies also show that HA modulates the activity of cholinergic neurons [58]. Moreover, it has been reported that thioperamide significantly improves the response latency in a passive avoidance response in senescence-accelerated mice [59, 60]. Thioperamide or clobenpropit administration in mice attenuates the amnesic effects of scopolamine in the elevated plus-maze test and the step-through passive-avoidance test and improves memory [61, 62].

## **Attention-Deficit Hyperactive Disorder (ADHD)**

 ADHD is an early childhood developmental disorder with underlying motor, emotional, attention and learning alterations. Abnormalities in monoamine neurotransmitters appear to contribute significantly to the disturbances in ADHD patients [63]. The use of  $H_3R$  antagonists can be useful in attention disorders on the basis of the vigilancepromoting effects of these drugs.  $H_3R$  antagonist treatments

have procognitive properties in animal models of learning and memory, as mentioned before in this review. In some studies, impairments in cognitive processes or motor patterns are similar to those observed in ADHD. In one of the study, administration of the selective  $H_3R$  antagonist GT2016 improved the rate of acquisition of a multi-trial passiveavoidance response [64, 65].

#### **H3R Ligands**

 $H_3R$  is an auto- and hetero- receptor, thus its activation reduces, whereas blockade increases, not only the release of histamine but also several other neurotransmitters.  $H_3R$  are not restricted to histaminergic neurons as histamine receptors are also found at various co-localized neurons [66].

 H3R agonists are mostly small molecules, having imidazole as nucleus and are derivatives of histamine. N-Methyl histamine, a potent agonist of histamine obtained by addition of methyl group at the basic amino group, is frequently used as radioligand. Imetit, immepip, impentamine, proxyfen, GT-2331 are some other H3R agonists [66, 67]. So far, efforts to replace the imidazole-moiety in agonists have been unsuccessful [68, 69]. Addition of the methyl group in imidazole side chain results in  $(R)$ - $\alpha$ -methyl histamine, which is a typical  $H_3R$  agonist, used for the early pharmacological characterization of the H3R [70]. Small structural changes lead to very potent ligands with selectivity for H3R receptors, like imetit or immepip. Methylation of the piperidine nitrogen of immepip gives methimepip, currently the most potent and selective  $H_3R$  agonist. Many  $H_3R$ ligands have been developed since then and are discussed below. Thioperamide was developed as first potent selective H3R antagonist and later clobenpropit was discovered as inverse agonist [71]. Ciproxyfan was also added to the antagonist series later [72]. The structures of the above compounds are given in (Fig. **3**).

 Impentamine, a higher homolog of histamine, proxyfan and cipralisant were originally characterized to be  $H_3R$ antagonists [73]. Thioperamide was the reference  $H_3R$ antagonist for almost two decades. Newer imidazole containing  $H_3R$  ligands as potent  $H_3R$  antagonists have also been discovered. These compounds have low bioavailability, poor blood-brain barrier penetration and CYP450 inhibition due to the imidazole-moiety. The replacement of the imidazole moiety is important towards more selective and drug-like  $H_3R$  antagonists [74, 75].

Initial pharmacophore modeling for  $H_3R$  antagonists was based on a set of imidazole antagonists and consisted of two hydrophobic pockets and four hydrogen binding sites, out of which two should be in imidazole ring [76]. Based on later studies, prototype  $H_3$  antagonist pharmacophore consisting of basic moiety separated by alkyl spacer to a central core was elucidated [77].

More recent studies shows that the  $H_3R$  antagonist pharmacophore consists of two protonation sites (i.e. basic centers) which are connected by central aromatic ring, alkyl



**Fig. (3).** Histamine and its early ligands.

spacer or hydrophobic region. The basic sites can simultaneously interact with Asp 114 and a Glu 206 which are believed to be the key residues that histamine interacts with to stabilize the receptor in the active state. The central region of these antagonists contains a lipophilic group, usually an aromatic ring that is believed to interact with several nearby hydrophobic side chains [78].

#### **New H3R Antagonists**

 The research of Mange and coworkers led to the development of potent H3R antagonist VUF-5391.James Black foundation discovered early nonimidazole  $H_3R$  antagonist, JB-98064. Aplysamine was first to be patented by Harbor Branch Oceanographic Institution containing phenoxyalkayl, N, N-dialkyalamine fragment. Ganellin and coworkers synthesized a new series of H3R antagonists (e.g. UCL-1972, UCL-2190, UCL-2173) for exploring the SAR of aplysamine [79, 80].

 Johnson and Johnson has contributed a lot in the development of  $H_3$  antagonists. A calcium channel blocker RWJ-20085 has been found to give high  $H_3R$  activity [81]. This compound has the N,N-dialkylamino propoxyphenyl motif of early compounds and also imidazopyrimidine motif. Optimization of RWJ-20085 led to the development of JNJ-6379490 which is having good  $H_3R$  antagonistic activity [82]. Another compound, JNJ-5207852 decreased the episodes of cataplexy in genetically narcoleptic Doberman Pinscher model. Unfortunately JNJ-5207852 showed tendency of phospholipidosis, so its development was stopped [83].

 Conformational restriction to aminoalkoxy side chain of RWJ-20085 provided JNJ-7737782 which was active in rodent model of wakefulness. Another molecule with rigidified linker, JNJ-10181457 has good  $H_3R$  selectivity, brain residency and pharmacokinetic profile and increases Ach and NE levels in rat frontal cortex [84]. JNJ-17216498 is a potent orally active  $H_3R$  antagonist that has completed phase-II trial for narcolepsy [85]. RWJ-333369 has also completed phase-II clinical trials for epilepsy [86]. Structures of some of the above compounds are given in (Fig. **4**).



**Fig. (4).** Structures of histamine and its ligands from Menge and coworkers, Harbor Branch Oceanographic Institution, James Black foundation, Gliatech, Johnson & Johnson and Bioproject.

 Gliatech investigated potential role of imidazole based compound GT-2331 having complex pharmacological profile. Some other compounds synthesized by Gliatech like GT-2394, GT-2227 and GT-2016 are in preclinical stage [87]. FUB UCL-INSERM Bioproject reported FUB-181, BF2.649, FUB-833 and FUB-836 as efficient and selective  $H_3$  antagonists with increased turnover of histamine, norepinephrine and serotonin in key brain regions [88, 89]. Replacement of imidazole with piperidine gave compound BF2.649 with increased ACh and brain dopamine level in rats [90-91]. Structures of Gliatech and Bioproject compounds are given in (Fig. **4**).

Abbott also developed numerous  $H_3R$  antagonists. A phenoxy propyl piperazine analogue, A-923, was having good antagonistic potency but its bioavailability profile was poor [92]. Further lead optimization of this compound led to the development of alanine type molecules A-304121 and A-317920. Compound A-317920 showed phospholipidosis, attributed to the dibasic amine and cationic amphiphillic nature of the compound [93, 94]. A bisphenyl motif was incorporated into some compounds such as A-331440 and

 To improve absorption and drug likeliness, ABT-239 was designed as conformationally restricted analogue with propyloxy spacer tied up in benzofuran. It has excellent selectivity for  $H_3R$  and also has good bioavailability in several species of rat, dog and monkeys. Inhibition of CYP 450 was also not observed. Increased acetylcholine, dopamine and histamine levels were reported in animal models after administration of same [97, 98]. It is presently in clinical development.

 Compounds A-688057 and A-687136 showed good affinity towards  $H_3R$ . Both of these compounds were effective in preclinical models of cognitive disorders. Lipophillicity was less as compared to ABT-239, so fewer incidences of phospholipidosis were observed. However, short half life due to rapid metabolism excluded the clinical development of both of these compounds [99, 100].



Fig. (5). New H<sub>3</sub>R antagonists from Abbott.

 Abott also synthesized naphthalene and quinoline based compounds having  $H_3R$  antagonistic profile [101]. Abott has also disclosed natural product connessine and its N- methyl D- alanine analogue as potent  $H_3$  antagonist [102].

 The quinoline compounds showed good *in vitro* potency, 90% oral bioavalability,  $t_{1/2}$  of 5.3 hrs and good brain penetration. The development was stopped because of photo instability of salts and cardiovascular side effects [103]. The structures of some compounds synthesized by Abbott are given in (Fig. **5**).

 Based on phenoxypropylamine compounds, GlaxoSmithkline have developed  $H_3R$  antagonists with tetra hydrobenzoazepine scaffold. Propyl piperidine motiff in initial compound GSK-1 could be replaced with various aryl and hetero aryl ethers. Compounds GSK-2 and GSK-207040, GSK-189254 are selective at  $H_3R$  with good brain penetration and also have good pharmacokinetic profiles [104, 105]. These compounds promote the release of acetylcholine, noradrenaline and dopamine in rat cortex [106]. Compound GSK-207040 entered in phase I trial for dementia and II for narcolepsy. GSK-239512 is also in clinical trials [107]. Some other GSK compounds, for pain and cognition activity, based on pyrazine and tetrahydrobenzoazepine motif are GSK-207040 and GSK-

334429 [108]. Both these compounds are inverse agonists of H3R. GSK-207040 has also been used for schizophrenia but not for positive symptoms. Structures of above compounds are given in (Fig. **6**).

 Merck/Banyu has come in focus with new series of potent CNS penetrating quinazolinones QNZ-1, QNZ-2 and QNZ-3 [109]. In this series, compound QNZ-3 showed a favorable pharmacokinetic profile without cardiovascular and CNS toxicity. The pyrrolidinylproyloxy side chain in compound QNZ-1 was constrained as a cyclobutylpiperidinyloxy moiety in compound QNZ-2 [110]. Banyu has also disclosed quinazolinone compound, QNZ-3 with favorable results. Another  $H_3$  antagonist MK0249 is reported to be in phase II clinical trials [111, 112]. Structures of these compounds are also given in (Fig. **6**).

 Pfizer compound PF-03654746 (Fig. **6**) is in phase I trial as  $H_3R$  antagonist. This compound enhanced the release of histamine in rat prefrontal cortex. It was developed from compound by optimizing the physiological properties to avoid phospolipidosis which was observed with early analogs. On the basis of favorable results, it entered the Phase II clinical trials but later discontinued due to insomia produced as side effect [113].



**Fig. (6).** New H3R antagonists from Glaxo Smithkline, Pfizer and Merck/Banyu.

Sr. No.	<b>Compound Name</b>	<b>Present Status</b>	<b>Use</b>	<b>References</b>
	BF2.649	Phase-III terminated	Narcolepsy, catalepsy	www.clinicaltrials.gov/23/12/2011
$\mathfrak{D}$	JNJ-17216498	Completed Phase-II	Narcolepsy	www.clinicaltrials.gov/23/12/2011
3	RWJ-333369	Phase-II completed	Epilepsy	www.clinicaltrials.gov/23/12/2011
$\overline{4}$	ABT-239	Phase-I completed	Obesity and cognitive disorders	94,95,96
5	GSK207040	Phase-I trial for dementia. Phase-II for narcolepsy	Dementia, narcolepsy	www.clinicaltrials.gov/23/12/2011
6	GSK-239512	Phase-I completed	Alzheimer	www.clinicaltrials.gov/23/12/2011
	PF-36504746	Phase-II completed	Schizophrenia, and ADHD	www.clinicaltrials.gov/23/12/2011

**Table 1. Current H3R Antagonists with their Present Status** 

#### **CONCLUSION**

 Brain histamine plays an important role in various CNS disorders. Histamine receptor antagonists cause the release of various neurotransmitters such as dopamine, gamma amino butyric acid, serotonin etc. Therefore, their role in epilepsy, schizophrenia, ADHD, narcolepsy and other central nervous system disorders is being explored. Few compounds that reached clinical trials were withdrawn due to phospholipidosis or pharmacokinetic constraints. Researchers are using the knowledge of these failures to prepare newer, selective  $H_3R$  antagonists with lesser side effects.

Current status of various representative  $H_3R$  ligands from different companies is given in Table **1**. Thus we can conclude that there is wide scope for the development of histamine H<sub>3</sub> receptor antagonists for the treatment of all above mentioned disorders.

# **CONFLICT OF INTEREST**

 The authors confirm that this article content has no conflicts of interest.

# **ACKNOWLEDGEMENTS**

Declared none.

### **REFERENCES**

- [1] Arrang, J.M.; Garbarg, M.; Schwartz, J.C. Auto inhibition of brain histamine release mediated by a novel class  $(H_3)$  of histamine receptor. *Nature,* **1983**, *302*, 832-837.
- [2] Brown, R.E.; Stevens, D.R.; Hass, H.L. The physiology of brain histamine. *Prog. Neurobiol*., **2001**, *63*, 637-672.
- [3] Hass, H.; Panula, P. The role of histamine and tuberomamillary nucleus in the central nervous system. *Nat. Rev. Neurosci.,* **2003**, *4*, 121-130.
- [4] Kazumori, H.; Ishihara, S.; Rumi, M.A.; Ortegacava, C.F.; Kadowaki,Y.; Kinoshita Y. Transforming growth factor- alpha directly augments histidine decarboxylase and vesicular monoamine transporter 2production in rat enterochromaffin like cells. *Am. J. Physiol. Gastrointest. Liver Physiol.,* **2004**, *286*, 508-514.
- [5] Zeldis, J.B.; Friedman, L.S.; Isselbacher, K.J. Ranitidine: A new H<sub>2</sub> receptor antagonist. *New Eng. J. Med.,* **1983**, *309*, 1368-1373.
- [6] Kerstin, S.; Tim, C.; Holger, S. Histamine H<sub>3</sub> receptor antagonists go to clinics. *Biol. Pharm. Bull.,* **2008**, *12*, 2163-2181.
- [7] Morisset S.; Rouleau, A.; Ligneau, X.; Gbahou, F.; Tardivel-Lacombe, J.; Stark, H.; Schunack, W.; Ganellin, CR.; Schwartz,

JW.; JM. High constitutive activity of native H-3 receptors regulates histamine neurons in brain. *Nature*, **2000**, *408*, 860-864.

- [8] Michael, B.; Christopher, W.B.; Manuel, de L.R. Histamine H<sub>3</sub> receptors as drug discovery target. *J. Med. Chem.*, **2011**, *54*,1-27.
- [9] Yokoyama, H.; Linuma, K. Studies and functional role of histaminergic neuron system by using pharmacological agents, knockout mice and positron emission tomography. *CNS Drugs,*  **1996**, *5*, 321-330.
- [10] Hass, H.L.; Sergeeva, O.A.; Selbach, O. Histamine in nervous system. *Physiol. Rev.,* **2008**, *88*, 1183-1241.
- [11] Lovenberg, T.W.; Roland, B.L.; Wilson, S.J.; Jiang x., Pyati, J.; Huvar, A.; Jackson, M. R.; Erlender, M.G. clonning and functional expression of human histamine H3 receptor. *Mol. Pharmacol.,* **1999**, *55*, 1101-1107.
- [12] Hancock, A.A.; Esbenshade, T.A.; Kruger, K.M.; Yao, B.B. Genetic and pharmacological aspects of histamine H<sub>3</sub> receptor heterogenecity. *Life Sci.,* **2003**, *55*, 1101-1107.
- [13] Bongers, G.; Bakker, R.A.; Leurs, R. Molecular aspectes of histamine H3 receptor. *Biochem. Pharmcol.,* **2007**, *73*, 1195-1204.
- [14] Lovenberg, T.W.; Pyati, J.; Chang, S.; Wilson S.J., Erlander M.G., Clonning of rat histamine  $H_3$  receptor reveals distinct species pharmacological profiles*. J. Pharmacol. Exp. Ther.,* **2000**, *293*, 1177-1178.
- [15] Gomez-Ramirez, J.; Ortiz, J., Blanco L. Presynaptic H3 autoreceptor modulate histamine synthesis through cAMP pathway. *Mol. Pharmacol*., **2002**, *61*, 239-245.
- [16] Churchill, J.A.; Gammon, G.D. The effect of antihistaminic drugs on convulsive seizure. *Am. Med. Assoc.,* **1949**, *141*, 18-21.
- [17] Tuomisto L.; Tacke U. 2-thiozolylethylamine, a seelective histamine H1 agonist decrease seizure susceptibility in mice. *Neuropharmacology,* **1986**, *25*, 955-958.
- [18] David, S. Molecular analysis of the histamine  $H_3$  Receptors. Ph.D. Thesis, The University of Regenburg: Germany, December **2009**.
- [19] Strasser, A.; Wittman, H. J.; Seifert, R. Ligand specific contribution of N terminus and E-2 loop to pharmacological properties of histamine H1 receptor. *J. Pharmacol. Exp. Ther.,* **2008**, *326*, 783-791.
- [20] Midzyanovskaya, I. S.; Tuomisto, L. Increased density of H1 histamine receptors in brain regions of rats with absence epilepsy. *Inflamm. Res.,* **2003**, *52*, S29-S30.
- [21] Yokoyama, H; Iinuma, K; Yanai, K; Watanabe, T; Sakurai, E; Onodera, K. Proconvulsant effect of ketotifen, a histamine H1antagonist, confirmed by the use of D-chlorpheniramine with monitoring electroencephalography. *Meth. Find Exp. Clin. Pharmacol*., **1993**, *15*, 183-188.
- [22] Vohora, D.; Pal, S.N.; Pillai, K.K. Histamine as an anticonvulsant inhibitory neurotransmitter. *Curr. Neuropharmacol*., **2004**, *2*, 419-425.
- [23] Yokoyama, H.; Onodera, K.; Iinuma, K.; Watanabe, T. Effect of thioperamide, a histamine  $H_3$  receptor antagonist, on electrically induced convulsions in mice. *Eur. J. Pharmacol*. **1993**, *234*, 129-133.
- [24] Witkin, J.M.; Nelson, D.L. Selective histamine H<sub>3</sub> receptor antagonists for treatment of cognitive deficiencies and other

disorders of the central nervous system. *Pharmacol. Ther.,* **2004**, *103*, 1-20.

- [25] Holger, S.; Bassem S.; Michel, K.; Annette, H.; Xavier, L.; Robill, G. Novel histamine  $H_3$  antagonist with carbonyl substituted 4-(3-(phenoxy)propyl)-1H-imidazole structures like ciproxifan and related compound. *J. Med. Chem*., **2000**, *43*, 3987-3994.
- [26] Vohora, D.; Pal, S.N.; Pillai, K.K. Histaminergic H<sub>3</sub>-receptors as modulators of CNS function. *Ind. J. Pharmacol.,* **2001**, *33*, 17-28.
- [27] Jia, F.; Kato, M.; Dai, H.; Xu, A.; Okuda, T.; Sakurai, E.; Okamura, N.; Lovenberg, T.W.; Barbier, A.; Carruthers, N.I.; Iinuma, K.; Yanai, K. Effects of histamine H<sub>3</sub> antagonists and donepezil on learning and mnemonic deficits induced by pentylenetetrazol kindling in weanling mice. *Neuropharmacology,* **2006**, *50*, 404-411.
- [28] Arias Montno, J.A.; Floran, B.; Gracia, M.; Aceves, J.; Young, J.M. Histamine H<sub>3</sub> receptor mediated inhibition of depolarization induced, dopamine  $D_1$  receptor dependant release of g-aminobutyric acid from rat striatal slices. *Br. J. Pharmacol.,* **2001**, *133*, 165-171.
- [29] Korotkova, T.M.; Hass, H.L.; Brown, R.E., Histamine Excites GABAergic cells in rat substantia nigra and ventral tegmental area *in vitro*. *Neurosci. Lett.,* **2002**, *320*, 133-136.
- [30] Bergquist, F.; Rut, J.A.; Hven, A.; Ludwig, M.; Dutia, M.B. Histaminergic and glycinergic modulation of GABA release in the vestibular in the nuclei of normal and labyrinthectomised rats. *J. Physiol.,* **2006**, *577*, 857-868.
- [31] Mclaughlin, F. L. Actylcholine in treatment of epilepsy. *Br. Med. J.*, **1933**, 1, 997-998.
- [32] Passani, M.B.; Cangioli, I.; Baldi, E.; Bucherelli, C.; Mannaioni, P.F.; Blandina, P. Histamine H<sub>3</sub> receptor-mediated impairment of contextual fear conditioning, and in-vivo inhibition of cholinergic transmission in the rat basolateral amygdala. *Eur. J. Neurosci.,*  **2001**, *14*, 1522-1532.
- [33] Cecchi, M.; Giorgetti, M.; Bacciottini, L.; Giovannini, M. G.; Blandina, P. The role of Interactions between histaminergic and cholinergic systems in learning and memory. *Inflamm. Res.,* **1998**, *47*, S32-S33.
- [34] Philippu, A.; Prast, H. Importance of histamine in modulatory processes, locomotion and memory. *Behav. Brain Res.,* **2001**, *124*, 151-159.
- [35] Kraus, M. M.; Fischer, H.; Tran, M. H.; Philippu, A.; Prast H., Modulation of acetylcholine release by histamine in the nucleus accumbens. *Inflamm. Res.*, **2001**, *50*, S74-S75.
- [36] Fink, K.; Schlicker, E.; Neise, A.; Gothert, M. Involvement of Presynaptic receptors in inhibitor effect of histamine on serotonin release in rat brain cortex. *Arch. Pharmacol.*, **2008**, *342*, 513-519.
- [37] Threlfell, S.; Cragg, S.J.; Kallo, I.; Turi, G.F.; Coen, C.W.; Greenfield, S.A., histamine H<sub>3</sub> receptor inhibits serotonin release in substantia nigra pars reticulate. *J. Neurosci.*, **2004**, *24*, 8704-8710.
- [38] Tokita, S.; Takahashi, K.; Kotani, H. Recent advances in molecular pharmacology of the histamine systems: physiology and pharmacology of histamine H3 receptor: roles in feeding regulation and therapeutic potential for metabolic disorders. *J. Pharmacol. Sci.*, **2006**, *101*, 12-18.
- [39] Masaki, T.; Yoshimatsu H., Neuronal histamine and its receptors in obesity and diabetes *Curr. Diabetes Rev.*, **2007**, *3*, 212-216.
- [40] Pollard, H.; Moreau, J.; Arrang, J.M; Schwartz, J.C. A detailed autoradiographic mapping of histamine H<sub>3</sub> receptors in rat brain areas. *Neuroscience,* **1993**, *52*, 169-189.
- [41] Masaki, T.; Yoshimatsu, H.; Chiba, S.; Watanabe, T.; Sakata, T. Central infusion of histamine reduces fat accumulation and upregulates UCP family in leptin resistant obese mice. *Diabetes*, **2001**, *50*, 376-384.
- [42] Matsui-Sakata, A.; Ohtani, H.; Sawada, Y. Receptor occupancy based analysis of the contributions of various receptors to antipsychotics-induced weight gain and diabetes mellitus. *Drug Metab. Pharmacokinet*., **2005**, *20*, 368-378.
- [43] Orgensen, E.A.; Knigge, U.; Warberg, J.; Kjaer, A. Histamine and the regulation of body weight. *Neuroendocrinology,* **2007**, *86*, 210-214.
- [44] Masaki, T.; Yoshimatsu, H.; The hypothalamic  $H_1$  receptor: a novel therapeutic target for disrupting diurnal feeding rhythm and obesity. *Trends Pharmacol. Sci.,* **2006**, *27*, 279-284.
- [45] Machidori, H; Sakata, T; Yoshimatsu, H.; Wada, H. Zucker obese rats: defect in brain histamine control on feeding. *Brain Res.,* **1992**, *590*, 180-186.
- [46] Monti, J. M.; Jantos, H.; Boussard, M.; Allier, H.; Orellana, C.; Olivera, **S.** Effects of selective activation or blockade of the

histamine H<sub>3</sub> receptor on sleep and wakefulness. *Eur. J. Pharmacol*., **1991**, *205*, 283-287.

- [47] Jones, B. E. From waking to sleeping: neuronal and chemical substrates. *Trends Pharmacol. Sci.*, **2005**, *26*, 578-586.
- [48] Parmentier, R.; Ohtsu, H.; Djebbara-Hannas, Z.; Valatx, J.L.; Watanabe, T.; Lin, J.S. Anatomical, physiological, and pharmacological characteristics of histidine decarboxylase knock-out mice: evidence for the role of brain histamine in behavioral and sleep-wake control. *J. Neurosci.,* **2002**, *22*, 7695-7711.
- [49] McLeod, R.L.; Aslanian, R.; Del Prado, M.; Duffy, R.; Egan, R.W.; Kreutner, W.; McQuade, R.; Hey, J. A. Sch 50971, An orally active histamine  $H_3$  receptor agonist, inhibits central neurogenic vascular inflammation and produces sedation in the guinea pig. *J. Pharmacol. Exp. Ther.*, **1998**, *287*, 43-50.
- [50] Kiyono, S.; Seo, M.L.; Shibagaki, M; Watanabe, T.; Maeyama, K.; Wada, H. Effects of alpha-fluoromethyl histidine on sleep-waking parameters in rats. *Physiol. Behav.*, **1985**, *34*, 615-617.
- [51] Lamberty,Y.; Margineanu, D.G.; Dassesse, D.; Klitgaard, H. H3 agonist immepip markedly reduces cortical histamine release, but only weakly promotes sleep in the rat. *Pharmacol. Res.,* **2003**, *48*, 193-198.
- [52] Le, S.; Gruner, J.A.; Mathiasen, J.R.; Marino, M.J.; Schaffhauser, H. Correlation between ex vivo receptor occupancy and wakepromoting activity of selective H3 receptor antagonists. *J. Pharmacol. Exp. Ther.,* **2007**, *325*, 902-909.
- [53] Mercier, V.G.; Sakai, K.; Jouvet, M. Specific neurons for wakefulness in the posterior hypothalamus in the cat. *C. R. Acad. Sci.,* **1984**, *298*,195-200.
- [54] Mochizuki, T.; Okakura-Mochizuki, K.; Horii, A.; Yamamoto, Y.; Yamatodani, A. Histaminergic modulation of hippocampal acetylcholine release *in vivo*. *J. Neurochem.,* **1994**, *62*, 2275-2282.
- [55] Haas, H; Panula, P. The role of histamine and the tuberomamillary nucleus in the nervous system. *Nat. Rev. Neurosci.,* **2003**, *4*, 121-130.
- [56] Panula, P.; Rinne, J.; Kuokkanen, K.; Eriksson, K.S.; Sallmen, T.; Kalimo, H.; Relja, M. Neuronal histamine deficit in Alzheimer's disease. *Neuroscience*, **1998**, *82*, 993-7.
- [57] Chiyomi, T.; Yukio, S.; Miyuki, N.; Yoko, F.; Chiaki, K. Effects of vasopressin on histamine  $H_1$  receptor antagonist-induced spatial memory deficits in rats. *Eur. J. Pharmacol.,* **2001**, *423*, 167-170.
- [58] Mochizuki, T.; Okakura, M.K.; Horii, A.; Yamamoto, Y.; Yamatodoni, A. Histaminergic modulation of hippocampal acetylcholine release *in vivo*. *J. Neurochem.,* **1994**, *62*, 2275 -2282.
- [59] Meguro, K.; Yanai, K.; Saaki, N.; Sakurai, E.; Maeyama, K.; Sasaki, H.; Watanabe, T. Effects of thioperamide, a histamine H<sub>3</sub> antagonist, on the step through passive avoidance response and histidine decarboxylase activity in senescence accelerated mice*. Pharmacol. Biochem. Behav.,* **1995**, *50*, 321-325.
- [60] Flood, J.F.; Uezu, K.; Morley, J.E. Effect of histamine  $H_2$  and  $H_3$ receptor modulation in the septum on post-training memory processing. *Psychopharmacology,* **1998**, *140*, 279-84.
- [61] Miyazaki, S.; Imaijumi, M.; Onodera, K. Effects of thioperamide on the cholinergic system and the step-through passive avoidance test in mice. *Methods Find. Exp. Clin. Pharmacol.,* **1995**, *17*, 653-658.
- [62] Orsetti, M.; Ghi, P.; Di Carlo, G. Histamine H(3)-receptor antagonism improves memory retention and reverses the cognitive deficit induced by scopolamine in a two-trial place recognition task. *Brain Res.,* **2001**, *124*, 235-242.
- [63] Oades, R.D. Attention deficit disorder with hyperactivity (ADDH): contribution of catecholaminergic activity. *Prog. Neurobiol.,* **1987**, *29*, 365-391.
- [64] Dumery, V.; Blozovski, D. Development of amygdaloid cholinergic mediation of passive avoidance learning in the rat., *Exp. Brain Res.,* **1987***, 67,* 61-69.
- [65] Biederman, J.; Spencer, T. Attention-deficit hyperactivity disorder (ADHD) as a noradrenergic disorder. *Biol. Psychiatry.,* **1999**, *46*, 1234-1242.
- [66] Leurs, R.; Vollinga, R.C.; Timmerman, H. Medicinal chemistry and theraputic potential of ligands of histamine H-3 receptors. *Prog. Drug. Res.,* **1995**, *45*, 107-165.
- [67] Stark, H.; Schilcker, E.; Schunack, W. Development of histamine H3 antagonist. *Drugs Future,* **1996**, *21*, 507-520.
- [68] De, Esch I.J.; Beljar, K.J. H<sub>3</sub> receptor agonists. *Mini Rev. Med. Chem.*, **2004**, *4*, 955-963.
- [69] Sylvain, C.; Maikel, W.; Patrice, T.; Rob, L.; Iwan, J.P.E. Histamine H3 receptor antagonist reach out for the clinics. *Drug Disc. Today*, *2005*, *10*, 1613-1627.
- [70] Meier, G.; Apelt, J.; Reichert, U.; Grabmann, S.; Ligneau, X.; Elz, S.; Leurquin, F.; Ganellin, C.R.; Schwartz, J. C.; Schunack, W.; Stark, H.; Influence of imidazole replacement in different structural classes of histamine H3-receptor antagonists., *Eur. J. Pharm. Sci.*, **2001**, *13*, 249-259.
- [71] Garbarg, M.; Arrang, J.M.; Rouleau, A.; Ligneau, X.; Dam, T.T.M.; Schwartz, J.C.; Ganellin, C.R. S-{2-(Imidazolyl)ethyl}isothiourea, A highly specific and potent H<sub>3</sub> receptor agonist., *J. Pharmacol. Exp. Ther.,* **1992**, *263*, 304-310.
- [72] Ligneau, X.; Lin, J.S.; Vani-Mercier, G.; Jouvet, M.; Muir, J.L.; Ganellin,C.R.; Stark, H.; Elz, S.; Schunack, W.; Schwartz, J. Neurochemical and behavioural effect of ciproxifan, a potent histamine H<sub>3</sub> receptor antagonist., *J. Pharmacol. Exp. Ther.*, **1998**, *287*, 658-666
- [73] Arrang, J.M.; Garbarg, M.; Lancelot, J.C.; Lecomte, J.M.; Pollard, H.; Robba, M.; Schunack, W.; Schwartz, J. C. Highly potent and selective ligands for histamine receptors. *Nature*, **1987**, *327*, 117-123.
- [74] Vander goot, H.; Schepers, M.J.P.; Sterk, G.J.; Timmerman, H. Isothiourea analogs of histamine as potent agonists or antagonists of histamine H3 receptor. *Eur. J. Med. Chem.,* **1992**, *27*, 511-517.
- [75] Kerstin, S.; Tim, K.; Holger, S. Histamine H<sub>3</sub> receptor antagonist go to the clinics. *Biol. Pharm. Bull.,* **2008**, *12*, 2163-2181.
- [76] De Esch, I.; Mills, J.; Perkins, T.; Romeo, G.; Hoffmann, M.; Wieland, K.; Leurs, R.; Menge, W.; Nederkoorn, P.; Dean, P.; Timmerman, H. Development of a pharmacophore model for histamine H<sub>3</sub> receptor antagonists, using the newly developed molecular modeling program SLATE. *J. Med. Chem.* **2001**, *44*, 1666-1674.
- [77] Celanire, S.; Wijtmans, M.; Talaga, P.; Leurs, R.; de Esch, I. J. P. Histamine H3 receptor antagonists reach out for the clinic. *Drug Disc. Today,* **2005**, *10*, 1613-1627.
- [78] Frank U. A.; Scott D.; Sandor S. Three-dimensional models of histamine H3 receptor antagonist complexes and their pharmacophore. *J. Mol. Gr. Mod., 2006, 24*, 456-464.
- [79] Cowart, M.; Altenbach, R.; Black, L.; Faghih, R.; Zhao, C.; Hancock, A.A. Medicinal chemistry and biological properties of non-imidazole histamine H3 antagonists. *Mini Rev. Med. Chem.,* **2004**, *4*, 979-992.
- [80] Leurs, R.; Bakker, R.A.; Timmerman, H.; De Esch, I. J. P. The histamine  $H_3$  receptor: From gene cloning to  $H_3$  receptor drugs. *Nat. Rev. Drug Discovery,* **2005**, *4*, 107-120.
- [81] Shah, C.; McAtee, L.; Breitenbucher, J.G.; Rudolph, D.; Li, X.; Lovenberg, T.W.; Mazur, C.; Wilson, S.J.; Carruthers, N.I. Novel human histamine H3 receptor antagonists. *Bioorg. Med. Chem. Lett.,* **2002**, *12*, 3309-3312.
- [82] Bonaventure, P.; Letavic, M.; Dugovic, C.; Wilson, S.; Aluisio, L. Pudiak, C.; Lord, B.; Mazur, C.; Kamme, F.; Nishino, S.; Carruthers, N.; Lovenberg, T. Histamine H<sub>3</sub> receptor antagonists: from target identification to drug leads. *Biochem. Pharmacol*., **2007**, *73*, 1084-1096.
- [83] Barbier, A.J.; Berridge, C.; Dugovic, C.; Laposky, A.D.; Wilson, S.J.; Boggs, J.; Aluisio, L.; Lord, B.; Mazur, C.; Pudiak, C. M.; Langlois, X.; Xiao, W.; Apodaca, R.; Carruthers, N.I.; Lovenberg, T.W. Acute wakepromoting actions of JNJ-5207852, a novel, diamine-based H3 antagonist. *Br. J. Pharmacol*., **2004**, *143*, 649-661.
- [84] Galici, R.; Boggs, J.D.; Aluisio, L.; Fraser, I.C.; Bonaventure, P.; Lord, B.; Lovenberg, T.W. JNJ-10181457, a selective nonimidazole histamine H3 receptor antagonist, normalizes acetylcholine neurotransmission and has efficacy in translational rat models of cognition. *Neuropharmacology*, **2009**, *56*, 1131-1137.
- [85] A safety and effectiveness study of a single dose of JNJ-17216498 in patients with narcolepsy. http://www.clinicaltrials.gov/ct2/show/ NCT00424931?intr="JNJ-17216498"&rank=1 (Accessed December 18, **2011**)
- [86] Safety and Efficacy of RWJ-333369 in the Treatment of Partial Epilepsy. http://www.clinicaltrials.gov/ct2/show/NCT00228969? term=RWJ&rank=14 (Accessed December 19, **2011**)
- [87] Tedford, C.E.; Yates, S.L.; Pawlowski, G.P.; Nalwalk, J.W.; Hough, L.B.; Amin, K. M.; Phillips, J.G.; Durant, G.J.; Frederickson, R.C.A. Pharmacological characterization of GT-2016, a non-thiourea-containing histamine H3 receptor antagonist: *in vitro* and *in vivo* studies. *J. Pharmacol. Exp. Ther.,* **1995**, *275*, 598-604.
- [88] Stark, H. Histamine  $H_3$ -receptor agonists and antagonists: chemical, pharmacological, and clinical aspects. *Chem. Mol. Aspects Drug Des. Action,* **2008**, *199*, 199-214.
- [89] Stark, H; Huls, A; Ligneau, X; Purand, K.; Pertz H; Arrang J. M.; Schwartz, J.C.; Schunack, W. Development of FUB 181, a selective histamine H3-receptor antagonist of high oral *in vivo* potency with 4- (omega-(arylalkyloxy)alkyl)-1H-imidazole structure., *Arch. Pharm.,* **1998**, *331*, 211-218.
- [90] Wijtmans, M.; Leurs, R.; de Esch, I. Histamine H<sub>3</sub> receptor ligands break ground in a remarkable plethora of therapeutic areas. *Expert Opin. Invest. Drugs,* **2007**, *16*, 967-985.
- [91] Ligneau, X.; Perrin, D.; Landais, L.; Camelin, J.C.; Calmels, T.P.G.; Berrebi-Bertrand, I.; Lecomte, J.M.; Parmentier, R.; Anaclet, C.; Lin, J.S.; Bertaina-Anglade, V.; la Rochelle, C. Drieu; d'Aniello, F.; Rouleau, A.; Gbahou, F.; Arrang, J.M.; Ganellin, C.R.; Stark, H.; Schunack, W.; Schwartz, J.C. BF2.649 [1-{3-[3-(4-chlorophenyl) propoxy] propyl} piperidine hydrochloride], a nonimidazole inverse  $a$ gonist/antagonist at the human histamine  $H_3$  receptor: Preclinical pharmacology. *J. Pharmacol. Exp. Ther.,* **2007**, *320*, 365-375.
- [92] Vasudevan, A.; Conner, S.E.; Gentles, R.G.; Faghih, R.; Liu, H.; Dwight,W.; Ireland, L.; Kang, C.H.; Esbenshade, T.A.; Bennani, Y.L.; Hancock, A. A. Synthesis and evaluation of potent pyrrolidine H3 antagonist. *Bioorg. Med. Chem. Lett*., **2002**, *12*, 3055-3058.
- [93] Apelt, J.; Ligneau, X.; Pertz, H.H.; Arrang, J.M.; Ganellin, C.R.; Schwartz, J.C.; Schunack, W.; Stark, H. Development of a new class of nonimidazole histamine  $H_3$  receptor ligands with combined inhibitory histamine N-methyltransferase activity. *J. Med. Chem.,*  **2002**, *45*, 1128-1141.
- [94] Fox, G.B.; Pan, J.B.; Radek, R.J.; Lewis, A.M.; Bitner, R.S.; Esbenshade, T.A.; Faghih, R.; Bennani, Y.L.; Williams, M.; Yao, B.B.; Decker, M.W.; Hancock, A.A. Two novel and selective nonimidazole H<sub>3</sub> receptor antagonists A-304121 and A-317920: II. *In vivo* behavioral and neurophysiological characterization. *J. Pharmacol. Exp. Ther.*, **2003**, *305*, 897-908.
- [95] Esbenshade, T.A.; Fox, G.B.; Krueger, K.M.; Baranowski, J.L.; Miller, T.R.; Kang, C.H.; Denny, L.I.; Witte, D.G.; Yao, B.B.; Pan, J.B.; Faghih, R.; Bennani, Y.L.; Williams, M.; Hancock, A.A. Pharmacological and behavioral properties of A-349821, a selective and potent human histamine  $H_3$  receptor antagonist. *Biochem. Pharmacol*., **2004**, *68*, 933-945.
- [96] Hancock, A.A.; Bennani, Y.L.; Bush, E.N.; Esbenshade, T.A.; Faghih, R.; Fox, G.B.; Jacobson, P.; Knourek-Segel, V.; Krueger, K.M.; Nuss, M.E.; Pan, J.B.; Shapiro, R.; Witte, D.G.; Yao, B.B. Antiobesity effects of A-331440, a novel nonimidazole histamine H3 receptor antagonist. *Eur. J. Pharmacol*., **2004**, *487*, 183-197.
- [97] Ku, Y.Y.; Pu, Y.M.; Grieme, T.; Sharma, P.; Bhatia, A.V.; Cowart, M. An efficient and convergent synthesis of the potent and selective H3 antagonist ABT-239. *Tetrahedron Lett.,* **2006**, *62*, 4584-4589.
- [98] Faghih, R.; Dwight, W.; Black, L.; Liu, H.; Gentles, R.; Phelan, K.; Esbenshade, T.A.; Ireland, L.; Miller, T.R.; Kang, C.H.; Krueger, K.M.; Fox, G.B.; Hancock, A.A.; Bennani, Y.L. Structure-activity relationships of non-imidazole  $H_3$  receptor ligands. Part Binding preference for D-amino acids motifs. *Bioorg. Med. Chem. Lett.,* **2002**, *12*, 2035-2037.
- [99] Cowart, M.; Faghih, R.; Curtis, M.P.; Gfesser, G.A.; Bennani, Y.L.; Black, L.A.; Pan, L.; Marsh, K.C.; Sullivan, J.P.; Esbenshade, T.A.; Fox, G.B.; Hancock, A.A. 4-(2-[2-(2(R)- Methylpyrrolidin-1-yl)ethyl]benzofuran-5-yl)benzonitrile and related 2-aminoethyl benzofuran  $H_3$  receptor antagonists potently enhance cognition and attention. *J. Med. Chem.* **2005**, *48*, 38-55.
- [100] Cowart, M.; Gfesser, G.A.; Browman, K.E.; Faghih, R.; Miller, T.R.; Milicic, I.; Baranowski, J.L.; Krueger, K.M.; Witte, D.G.; Molesky, A.L.; Komater, V.A.; Buckley, M.J.; Diaz, G.J.; Gagne, G.D.; Zhou, D.; Deng, X.; Pan, L.; Roberts, E.M.; Diehl, M.S.; Wetter, J.M.; Marsh, K.C.; Fox, G.B.; Brioni, J.D.; Esbenshade, T.A.; Hancock, A.A. Novel heterocyclic-substituted benzofuran histamine  $H_3$ receptor antagonists: *in vitro* properties, drug-likeness, and behavioral activity. *Biochem. Pharmacol.,* **2007**, *73*, 1243-1255.
- [101] Pu, Y.M.; Ku, Y.Y.; Grieme, T.; Black, L.A.; Bhatia, A.V.; Cowart, M. An expedient and multikilogram synthesis of a naphthalenoid  $H_3$ antagonist. *Org. Process Res. Dev.,* **2007**, *11*, 1004-1009.
- [102] Zhao, C.; Sun, M.; Bennani, Y.L.; Gopalakrishnan, S.M.; Witte, D.G.; Miller, T.R.; Krueger, K.M.; Browman, K.E.; Thiffault, C.; Wetter, J.; Marsh, K.C.; Hancock, A.A.; Esbenshade, T.A.; Cowart,

M. D. The alkaloid conessine and analogs as potent histamine  $H_3$ receptor antagonists*. J. Med. Chem.,* **2008**, *51*, 5423-5430.

- [103] Altenbach, R.J.; Liu, H.; Banfor, P.N.; Browman, K.E.; Fox, G.B.; Fryer, R.M.; Komater, V.A.; Krueger, K.M.; Marsh, K.; Miller, T.R.; Pan, J.B.; Pan, L.; Sun, M.; Thiffault, C.; Wetter, J.; Zhao, C.; Zhou, D.; Esbenshade, T.A.; Hancock, A.A.; Cowart, M.D. Synthesis, potency, and *in vivo* profiles of quinoline containing histamine H<sub>3</sub> receptor inverse agonists. *J. Med. Chem.*, **2007**, *50*, 5439-5448.
- [104] Hudkins, R.L.; Raddatz, R. Recent advances in drug discovery of histamine H3 antagonists. *Ann. Rep. Med. Chem.,* **2007**, *42*, 49-62.
- [105] Medhurst, A.D.; Atkins, A.R.; Beresford, I.J.; Brackenborough K.; Briggs, M.A.; Calver, A.R.; Cilia, J.; Cluderay, J.E.; Crook, B.; Davis, J.B.; Davis, R.K.; Davis, R.P.; Dawson, L.A.; Foley, A.G.; Gartlon, J.; Gonzalez, M.I.; Heslop, T.; Hirst, W.D.; Jennings, C.; Jones, D.N.C.; Lacroix, L.P.; Martyn, A.; Ociepka, S.; Ray, A.; Regan, C.M.; Roberts, J.C Schogger, J.; Southam, E.; Stean, T.O.; Trail, B.K.; Upton, N.; Wadsworth, G.; Wald, J.A.; White, T.; Witherington, J.; Woolley, M.L.; Worby, A.; Wilson, D.M. GSK189254, a novel H3 receptor antagonist that binds to histamine H3 receptors in Alzheimer's disease brain and improves cognitive performance in preclinical models. *J. Pharmacol. Exp. Ther.,* **2007**, *321*, 1032-1045.
- [106] Medhurst, A.D.; Briggs, M.A.; Bruton, G.; Calver, A.R.; Chessell, I.; Crook, B.; Davis, J.B.; Davis, R.P.; Foley, A.G.; Heslop, T.; Hirst, W.D.; Medhurst, S.J.; Ociepka, S.; Ray, A.; Regan, C.M.; Sargent, B.; Schogger, J.; Stean, T.O.; Trail, B.K.; Upton, N.; White, T.; Orlek, B.; Wilson, D.M. Structurally novel histamine H<sub>3</sub> receptor antagonists GSK207040 and GSK334429 improve scopolamine-induced memory impairment and capsaicin-induced secondary allodynia in rats. *Biochem. Pharmacol.,* **2007**, *73*, 1182- 1194.
- [107] Bridging Study with GSK239512 in patients with mild to moderate Alzheimer's disease. http://www.clinicaltrials.gov/ct2/

Received: February 06, 2012 **Revised: June 13, 2012** Accepted: June 18, 2012

show/NCT00675090?term=GSK-239512&rank=1 (Accessed December 19, **2011**)

- [108] Southam, E.; Cilia, J.; Gartlon, J.E.; Woolley, M.L.; Lacroix, L.P.; Jennings, C.A.; Cluderay, J.E.; Reavill, C.; Rourke, C.; Wilson, D.M.; Dawson, L. A.; Medhurst, A.D.; Jones, D. Preclinical investigations into the antipsychotic potential of the novel histamine  $H_3$  receptor antagonist GSK207040. *Psychopharmacology,* **2009**, *201*, 483-494.
- [109] Nagase, T.; Mizutani, T.; Ishikawa, S.; Sekino, E.O.; Sasaki, T.; Fujimura, T.; Ito, S.; Mitobe, Y.; Miyamoto, Y.; Yoshimoto, R.; Tanaka, T.; Ishihara, A.; Takenaga, N.; Tokita, S.; Fukami, T.; Sato, N. Synthesis, structure-activity relationships, and biological profiles of a quinazolinone class of histamine  $H_3$  receptor inverse agonists. *J. Med. Chem.,* **2008**, *51*, 4780-4789.
- [110] Nagase, T.; Mizutani, T.; Sekino, E.; Ishikawa, S.; Ito, S.; Mitobe, Y.; Miyamoto, Y.; Yoshimoto, R.; Tanaka, T.; Ishihara, A.; Takenaga, N.; Tokita, S.; Sato, N. Synthesis and evaluation of structurally constrained quinazolinone derivatives as potent and selective histamine H<sub>3</sub> receptor inverse agonists. *J. Med. Chem.*, **2008**, *51*, 6889-6901.
- [111] Nagase, T.; Sato, N.; Kii, S.; Sato, K.; Tsuritani, T.; Sawada, N. Process for Production of 4(3H)-Quinazolinone Derivative. *PCT Int. Appl*., **2006**, (E-pub ahead of print).
- [112] Mizutani, T.; Nagase, T.; Ito, S.; Miyamoto, Y.; Tanaka, T.; Takenaga, N.; Tokita, S.; Sato, N. Development of novel 2- [4- (aminoalkoxy)phenyl]-4(3H)-quinazolinone derivatives as potent and selective histamine H<sub>3</sub> receptor inverse agonists. *Bioorg. Med. Chem. Lett.,* **2008**, *18*, 6041-6045.
- [113] A Study to Evaluate the Efficacy and Safety of Two Doses of PF-03654746 in Adults with Attention Deficit Hyperactivity Disorder (ADHD). http://www.clinicaltrials.gov/ct2/show/ NCT00531752?term=PF-03654746&rank=1 (Accessed December 19, **2011**).